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Describing morphological characters of seedlings of some dicotyledonous weeds for their identification and management

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Article information ABSTRACT

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Key words

Dicotyledonous, Morphological, Seedlings, Weeds identification Seedling traits of 60 weeds under 24 families of Magnoliopsida have been studied in some crop fields of Dakshin Dinajpur district of West Bengal. Conservative seedling characters emphasize that the studied taxa may be enlisted in four artificial groups *i.e*, Type I to IV, each having some families and/ or genera. Artificial keys have been made for identification under field observations. The bearing of this study has also been addressed by camping seedling data with other botanical disciplines. Peculiar juvenile behaviors like heteroblastic developments have also been observed. Seedling study is very much significant for eradication of weeds at juvenile stage before display of variable weeds in crop fields.

INTRODUCTION

Weeds are undesired plant species that grow with cultivated crops and intervene or compete with the crops for growth and nutrients and in this way affect the productivity leading to economic loss (Marwart et al. 2013). Most of the weeds are annual and complete their life cycle within a short period of time producing a large number of viable seeds that germinates immediately in almost every season to interact with crops. Therefore, rapid and accurate identification of weeds in seedling stage might be helpful for a successful weed management that can save both time and cost of production as well as lower chemical herbicide usage (Parkinson et al. 2013).

Importance of studying phenotypic traits of weed seedlings for their correct identification has a major role in suggesting suitable post-emergence herbicides for effective weed management. It is a common practice for the farmers in most parts of Eastern India that they use herbicides on mature weeds during pre or post harvesting period while abundant seeds have already been added by the weeds in the soil. Seeds protected by hard seed coats are not affected by the herbicides and they germinate into the next generation of weeds demanding proportional use of herbicides over the years. Hence, a strategy of weed management using herbicide or manual labor may be administered at the seedling stage i.e., before flowering and fruiting to stop the next generation before it germinates. For this, weeds must be

identified at the seedling stage through proper keys constructed based of their juvenile traits of cotyledons or other parts, which are conservative and viable for weeds growing in any geological or ecological conditions and this process has already advocated by some workers (Parkinson *et al.* 2013, Chomas *et al.* 2001, Chancellor 1966).

MATERIALS AND METHODS

Thorough survey for collection of seeds and/or seedlings has been done from March, 2017 to February, 2018 in the crop fields located in different mouzas (administrative unit) of Balurghat block. The list of seedlings studied is given in Table 1. The taxa are arranged family wise after Takhtajan, 1997 and alphabetically with author's name(s), and photograph number within each family. The seedlings are collected in pre- and post-harvesting periods as well as growth stages of crops. The seeds are air-dried and sown in prepared seedbeds separately from time to time with proper tagging to raise seedlings in the experimental garden of Balurghat College. So-raised seedlings were compared to natural ones for proper identification. Few seedlings were also identified following literatures of Chancellor (1966), Chomas et al. (2001) and Parkinson et al. (2013). The seedlings were described with following Duke (1965), Burger (1972), de Vogel (1980), Paria et al. (1990, 2006), and Das and Kamilya (2014). Field photographs taken with Nikon digital camera of all the seedlings with highlighted paracotyledons and first two leaves have

Table 1. List of species studied with author's names and photograph numbers arranged under each families following Takhtajan (1997)

1akntajan (1997)	
Ranunculaceae	Oxalidaceae
Ranunculus sceleratus L. [Figure A1]	Oxalis corniculata L. [Figure B15]
Papaveraceae	Apiaceae
Argemone mexicana L. [Figure A2]	Centella asiatica Urban [Figure C1]
Fumaria indica (Hasskn.) Pugsley [Figure A3]	Hydrocotyle sibthorpioides Lam. [Figure C2]
Molluginaceae	Oenanthe benghalensis (Roxb.) Benth. & Hook. f. [Figure C3]
Glinus lotoides L. [Figure A4]	Asteraceae
Glinus oppositifolius (L.) Aug. DC. [Figure A5]	Ageratum conyzoides L. [Figure C4]
Caryophyllaceae	Ageratum haustonianum Mill. [Figure C5]
Drymaria cordata (L.) Willd. exSchult. [Figure A6]	Centipeda minima (L.) A.Br. & Asch. [Figure C6]
Polycaarpon prostratum (Forssk.) Asch. & Schweinf. [Figure A7]	Eclipta prostrata (L.) L. [Figure C7]
Portulacaceae	Gnaphalium polycaulon Pers. [Figure C8]
Portulaca oleracea L. [Figure A8]	Grangea maderaspatana (L.) Poir. [Figure C9]
Amaranthaceae	Xanthium strumarium L. [Figure C10]
Achyranthus aspera L. [Figure A9]	Rubiaceae
Alternanthera paronychoides A.St. Hil. [Figure A10]	Dentella repens L. [Figure C11]
Alternanthera sessilis (L.) R.Br. ex DC. [Figure A11]	Oldenlandia corymbosa L. [Figure C12]
Amaranthus gangeticus L. [Figure A12]	Convolvulaceae
Amaranthus viridis L. [Figure A13]	Evolvulus nummularius (L.) L. [Figure C13]
Chenopodiaceae	Solanaceae
Chenopodium album L. [Figure A14]	Physalis minima L. [Figure C14]
Chenopodium ambrosioides L. [Figure A15]	Physalis peruviana L. [Figure C15]
Polygonaceae	Solanum nigrum L. [Figure D1]
Persicaria hydropiper (L.) Delabre [Figure B1]	Nicotiana plumbaginifolia Viv. [Figure D2]
Persicaria orientalis (L.) Spach [Figure B2]	Boraginaceae
Polygonum plebeium L. [Figure B3]	Coldenia procumbens L. [Figure D3]
Rumex dentatus L. [Figure B4]	Heliotropium indicum L. [Figure D4]
Malvaceae	Scrophulariaceae
Sida rhomboidea Roxb. exFlaming [Figure B5]	Lindernia ciliata (Colsm.) Pennell [Figure D5]
Sterculiaceae	Lindernia crustacea (L.) F. Muell. [Figure D6]
Melochia corchorifolia L. [Figure B6]	Lindernia nummulariifolia (D. Don) Wettst. [Figure D7]
Urticaceae	Lindernia parviflora (Roxb.) Haines [Figure D8]
Pouzolzia zeylanica (L.) Benn. & R.Br. [Figure B7]	Lindernia procumbens (Krock.) Philcox [Figure D9]
Euphorbiaceae	Mazus pumilus (Burm. f.) Steinis [Figure D10]
Acalypha indica L. [Figure B8]	Mecardonia procumbens (Mill.) Small. [Figure D11]
Chrozophora rottleri (Geiseler) A. Juss. exSpreng. [Figure B9]	Scoparia dulcis L. [Figure D12]
Croton bonplandianus Baill. [Figure B10]	Acanthaceae
Euphorbia hirta L. [Figure B11]	Hygrophilla difformis Blume [Figure D13]
Onagraceae	Lamiaceae
Ludwigia perennis L. [Figure B12]	Leucas aspera (Willd.) Link. [Figure D14]
Fabaceae	Salvia plebeia R.Br. [Figure D15]
Desmodium triflorum (L.) DC. [Figure B13]	1 L 0 -J
Senna tora (L.) Roxb. [Figure B14]	
han diagland in Disks A to D. A told (Table 2) has	IZ. A. A. C. P. C. P CT I

been displayed in Plate A to D. A table (Table 2) has been prepared with the major qualitative traits (given in abbreviated form) used for the diagnosis of the seedlings. Artificial keys have been constructed using both qualitative and quantitative traits to identify the seedlings of the weeds. In the key to the families, single species in a family has been mentioned in the parenthesis.

Artificial key (applicable for the studied taxa only)

Key to the seedling types

1. First two leaves subopposite to alternate2
1a. First two leaves opposite3
2. First two leaves exstipulateType I
2a. First two leaves stipulateType II
3. First two leaves exstipulateType III
3a. First two leaves stipulateType IV

Ke	y to the families of Type I
1.	Seedlings with latex; venation of paracotyledons parallelodromous
1a.	Seedlings without latex; venation of paracotyledons hyphodromous or actinodromous
2.	First two leaves with three or more primary veins
2a.	First two leaves with single primary vein4
3.	Margin of second leaf entire
3a.	Margin of second leaf crenate
	Apiaceae
4.	Pracotyledons narrowly oblong, apex obtuse; first internode angular5
4a.	Paracotyledons otherwise, apex acute or rounded first internode round

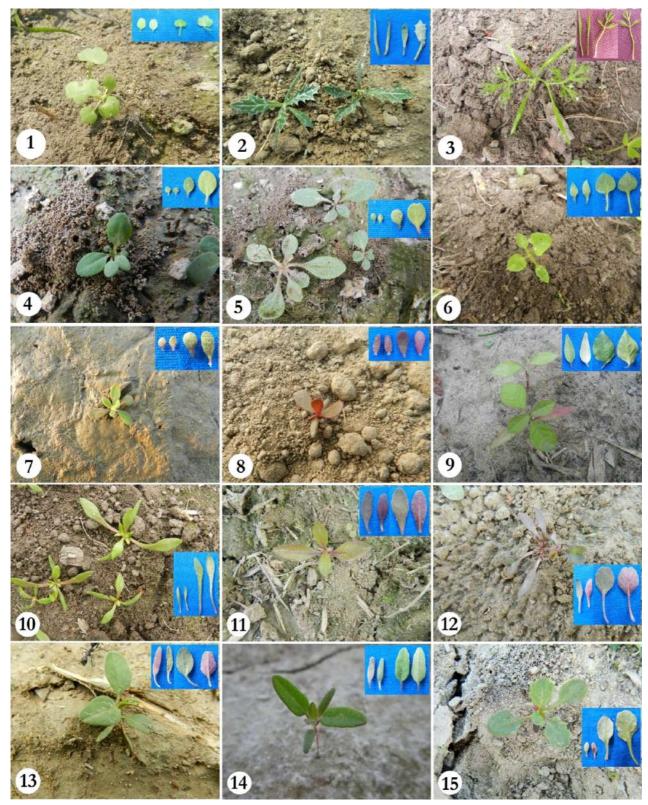


Plate A: 1. Ranunculus sceleratus; 2. Argemone mexicana; 3. Fumaria indica; 4. Glinus lotoides; 5. Glinus oppositifolius; 6. Drymaria cordata; 7. Polycarpon prostratum; 8. Portulaca oleracea; 9. Achyranthus aspera; 10. Alternanthera paronychioides; 11. Alternanthera sessilis; 12. Amaranthus gangeticus; 13. Amaranthus viridis; 14. Chenopodium album; 15. Chenopodium ambrosioides.

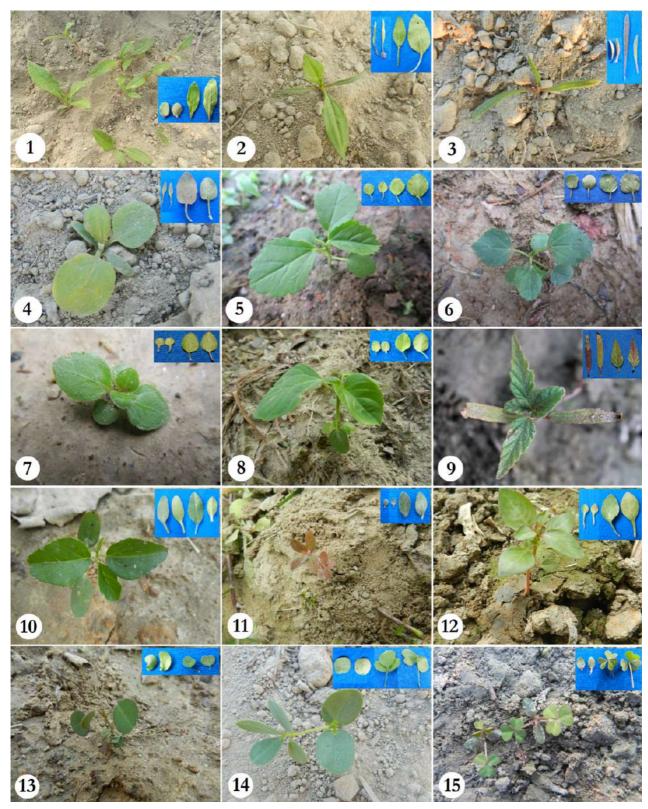


Plate B: 1. Persicaria hydropiper; 2. Persicaria orientalis; 3. Polygonum plebeium; 4. Rumex dentatus; 5. Sida rhomboidea; 6. Melochia corchorifolia; 7. Pouzolzia zeylanica; 8. Acalypha indica; 9. Chrozophora rottleri; 10. Croton bonplandianus; 11. Euphorbia hirta; 12. Ludwigia perennis; 13. Desmodium triflorum; 14. Senna tora; 15. Oxalis corniculata.

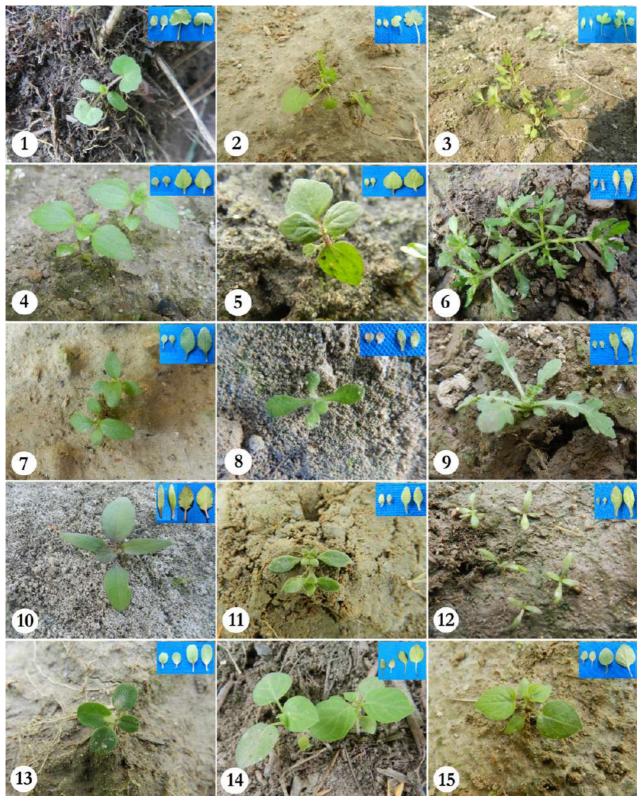


Plate C: 1. Centella asiatica; 2. Hydrocotyle sibthorpioides; 3. Oenanthe benghalensis; 4. Ageratum conyzoides; 5. Ageratum haustonianum; 6. Centipeda minima; 7. Eclipta prostrate; 8. Gnaphalium polycaulon; 9. Grangea maderaspatana; 10. Xanthium strumarium; 11. Dentella repens; 12. Oldenlandia corymbosa; 13. Evolvulus nummularius; 14. Physalis minima; 15. Physalis peruviana.

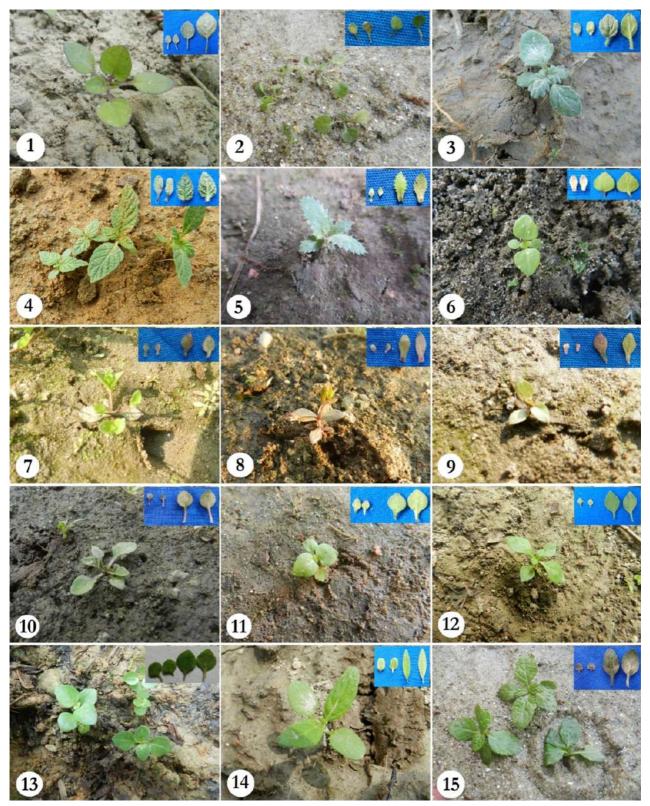


Plate D: 1. Solanum nigrum; 2. Nicotiana plumbaginifolia; 3. Coldenia procumbens; 4. Heliotropium indicum; 5. Lindernia ciliata; 6. Linternia crustacea; 7. Lindernia nummulariifolia; 8. Lindernia parviflora; 9. Lindernia procumbens; 10. Mazus pumilus; 11. Mecardonia procumbens; 12. Scoparia dulcis; 13. Hygrophilla difformis; 14. Leucas aspera; 15. Salvia plebeian.

	Нур	ocotyl				Paracoty	ledons											First two	leaves						First ernod
Name of the species	Shape	Surface	Texture	Surface	Petiole	Shape	Base	Apex	Margin	Pri. veins	Venation	hyllotaxy	Nature	Texture	Surface	Petiole	Stipule	Shape	Base	Apex	Margin	Pri.veins	Venation	Shape	c
Ranunculus sceleratus	r	glb	her	glb	1	obl	s.rnd	rnd	ent	1	hyp	alt	smp	her	glb	1	0	3lb	s.crd	rnd	ent	3	act	r	gl
Argemone mexicana	r	glb	thk	glb	0	lin	cnt	acu	ent	4	prl	alt	smp	her	glb	0	0	spt	atn	acu	spn-dnt	1	cam	r	gl
Fumaria indica	r	glb	her	glb	1	lin	atn	obt	ent			alt	smp	her	glb	1	0	pinn	cnt	acu	ent	1	cam	r	gl
Glinus lotoides	r	glb	her	glb	1	ov-s.orb	cnt	obt	ent	1	hyp	alt		her	glb	1	1	ov-elp	cnt	rnd	ent	1	hyp	an	gl
Glinus oppositifolius	r	glb	her	glb	1	obvs.orb	cnt	rnd	ent	1	hyp	alt		her	glb .	1	1	obv-elp	atn	rnd	ent	1	hyp	an	gl
Drymaria cordata	4-an	glb	her	glb	1	ov-lnc	cnt	acu	ent	1	hyp	opp		her		1	0	b.ov	s.trn	acu	ent	1	cam	4-an	
Polycaarpon prostratum		glb	her	glb	1	ov-elp	s.rnd	s.rnd	ent	1	hyp	opp	smp	her	glb	1	0	obv	ent	s.rnd	ent	1	hyp	4-an	toı
Portulaca oleracea Achyranthus aspera	r r	glb glb	fls her	glb glb	1	obl Inc	s.rnd ent	obt acu	ent	1	hyp	opp	smp	her	glb	1	0	obv elp-rhm	ent	s.rnd acu	ent ent	1	hyp	r 4-an	gl pu
Acnyraninus aspera Alternanthera	r	scb	fls	glb	1	n.lnc	atn	obt	ent	1	cam hyp	opp opp	smp smp	her her	pub sp.pub	1	0	oblnc	atn	s.acu	ent	1	cam	r r	pu
paronychioides Alternanthera sessilis	r	glb	fls	glb	1	elp-obl	atn	rnd	ent	1	hyp	орр		her	glb	1	0	obv	atn	obt	ent	1	cam	4-an	pu
Amaranthus gangeticus	r	glb	her	glb	1	n.obl	cnt	obt	ent	1	hyp	alt	F	her	glb	1	0	obv	ent	emr	ent	1	cam	4-an	Γ.
Amaranthus viridis	r	glb	her	glb	1	n.obl	cnt	obt	ent	1	hyp	alt		her	glb	1	0	ov-elp	ent	rts	ent	1	cam	4-an	
Chenopodium album	r	glb	fls	glb	1	n.obl	cnt	obt	ent	1	hyp	alt	smp	crs	scb	1	0	obl-lnc	cnt	obt	ent	1	cam	4-an	pi
Chenopodium ambrosioides	r	glb	her	glb	1	n.obl	cnt	obt	ent	1	hyp	alt	smp	crs	glb	1	0	ov-obl	cnt	s.rnd	s.ent	1	cam	4-an	
Persicaria hydropiper	r	glb	her	glb	1	obt-recs.orb		rnd	ent	1	cam	alt		cor	glb	1	1	elp-obl	atn	obt	ent	1	cam	r	sp.
Persicaria orientalis	r	glb	her	glb	1	lin	atn	acu	ent	1	hyp	alt	smp	her	pub	1	1	elp-lnc	atn	acu	ent	1	cam	r	h
Polygonum plebeium	r	scb	her	glb	0	lin-flc	atn	obt	ent	1	hyp	alt		her	glb	1	1	lin	atn	acu	ent	1	hyp	r	S
Rumex dentatus	r	glb	her	glb	1	n.obl	cnt	obt	ent	1	hyp	alt		her	glb	1	1	ov	s.trn	obt	ent	1	cam	r	g
Sida rhomboidea	r	hrs	her	hrs	1	s.orb	s.rnd	s.rts	ent	3	acr	alt		her	pub	1	f.1	ov.rhm	ent	obt	srt	3	act	r	h
Melochia corchorifolia	r	pub	her	glb	1	s.orb	rnd	rnd	ent	5	act	alt		her	pub	1	1	s.orb.b.ov	s.rnd	obt	crn-srt	3	act	r	p
Pouzolzia zeylanica	r	scb	her	d.pub	1	orb.ov	s.rnd	rts	ent	1	cam	opp	smp	her	hrs	1	1	ov	s.trn	obt	ent	3	act	4-an	m.
Icalypha indica	r	scb	her	sp.pub	1	elp-s.orb	rnd	s.trn	ent	3	acr	opp	smp	her	scb	1	1	ov	rnd	acu	srt	3	act	r	S
Chrozophora rottleri Croton bonplandianus	r	scb stl	her	sp.pub stl	1	b.elp.s.orb obl	rnd ent	s.trn s.rnd	ent	3	acr acr	opp alt		her her	scb stl	1	1	ov ov-elp	rnd s.rnd	acu acu	d.srt srt	3	act	r r	S
Suphorbia hirta	r	glb	her	glb	1	obl		s.rnd	ent	1	hyp	opp		her	pub	1	0	obv	ent	s.rnd	ent	1	cam	r	d.i
udwigia perennis	r	glb	her	glb	1	ov	b.cnt	acu	ent	i	hyp	opp	smp		glb	1	0	ov	b.ent	acu	ent	1	cam	r	g
Desmodium triflorum	r	pub	thk	scb	0	flc-obl	obq	rnd	ent	3	act	opp			glb	1	f.1	b.obv	b.cnt	sh.emr	ent	1	cam	6-an	h
Senna tora	r	sp.pub	thk	glb	0	obv-obl	aur	rnd	ent	3	act	alt			pub		f.1	obv	obq.trn	rts	ent	1	cam	4-an	p
Oxalis corneculata	r	sp.pub	her	glb	1	ov.elp	cnt	rnd	ent	1	hyp	alt			glb	1	1	oberd	cnt	dp.emr	ent	3	act	r	ĥ
Centella asiatica	r	glb	her	glb	1	obl-rec	s.rnd	rnd	ent	1	hyp	alt	smp	her	glb	1	0	s.ren	crd	rnd	crn	5	act	r	g
Hydrocotyle sibthorpioides	r	glb	her	glb	1	ov	s.rnd	rnd	ent	1	hyp	alt		her	glb	1	0	ren	s.crd	rnd	und	78		r	g
Denanthe benghalensis	r	glb	her	glb	1	elp	cnt	obt	ent	1	hyp	alt	smp		glb	1	0	3-lb	s.trn	rnd		3	act	r	g
Ageratum conyzoides	r	scb	her	glb	1	ov	cnt	rts	ent	1	hyp	opp	smp	her	m.pub	1	0	ov	s.trn	acu	d.srt	3	acr	r	p
Ageratum haustonianum Zentipeda minima	r r	scb glb	her her	scb glb	1	ov	cnt	rts s.rnd	ent	1	hyp	opp opp	smp smp	her	pub glb	1	0	ov elp-obv	b.cnt atn	obt obt	d.srt ent	3	acr hyp	r r	h g
Eclipta prostrata	r	glb	her	glb	0	elp obvs.orb	ent	rnd	ent	1	hyp	opp	smp	her	pub	1	0	ov-elp	ent	s.acu-obt	d.srt	3	acr	r	g p
Gnaphalium polycaulon	r	glb	her	glb	1	s.orb	s.rnd	rnd	ent	1	hyp	alt		her	d.pub	1	0	elp	atn	obt	ent	1	hyp	r	to
Grangea maderaspatana	r	glb	her	glb	0	obv-elp	atn	s.rnd	ent	1	hyp	alt		her	pub	1	0	obv-obl	atn	obt	ent	1	hyp	r	h
Canthium strumarium	r	glb	cor	glb	1	elp-lnc	atn	s.acu	ent	3	acr	opp	smp	cor	d.pub	1	0	ov	s.rnd	acu	srt	3	acr	r	h
Dentella repens	4-an	glb	her	glb	1	b.ov	rnd	obt	ent	1	hyp	opp	smp	her	glb	1	int	n.elp	cnt	acu	ent	1	hyp	4-an	g
Oldenlandia corymbosa	r	glb	her	glb	1	ov-s.orb	s.trn	rnd	ent	3	act	opp	smp		scb	1	1	n.elp	cnt	obt	ent	1	cam	4-an	g
Evolvulus nummularius	r	glb	her	glb	1	rec-s.orb	b.cnt	rnd		3	act	alt	smp		glb	1	0	obv	b.cnt	rts	ent	1	cam	r	h
Physalis minima		m.pub	her	scb,	1	ov	rnd	acu	ent	1	hyp	alt	smp		s.glb	1	0	ov	obq.s.rnd	acu	ent	1	cam	r	p
Physalis peruviana	r	pub	her	sp.pub	1	ov	b.cnt	acu	ent	1	hyp	alt			pub	1	0	ov	obq.s.rnd	obt	irr.und	1	cam	r	to
olanum nigrum Vicotiana plumbaginifolia	r r	pub glb	her her	glb glb	1	ov-lnc s.orb	ent s.rnd	acu rnd	ent ent	1	hyp hyp	alt alt		her her	m.pub glb	1	0	b.ov b.ov	s.rnd s.trn	acu acu	ent	1	cam cam	r r	h g
Coldenia procumbens	r	d.pub	cor	hrs	1	ov.rec	b.cnt	rnd	ent	1	hyp	opp	smp	her	vil	1	0	ov	b.cnt	acu	ent	1	cam	r	g
Heliotropium indicum	r	pub	her	pub	1	ov-obl	cnt	rnd	ent	1	hyp	opp		her	d.pub	1	0	ov	rnd	acu	ent	1	cam	r	h
Lindernia ciliata	r	glb	her	scb	0	ov	cnt	obt	ent	1	hyp	opp	smp	her	hrs	0	0	obl-spat	atn	acu	dnt	1	hyp	4-an	
indernia crustacea	r	scb	$\pm thk \\$	glb	1	ov	s.trn	obt	ent	1	hyp	opp	smp	her	glb	1	0	ov	s.trn	obt	crn	1	hyp	4-an	g
indernia nummulariifolia	r	glb	her	glb	1	elp	cnt	acu	ent	1	hyp	opp	smp	her	glb	1	0	ov-elp	cnt	acu	ent	1	hyp	4-an	g
indernia parviflora	r	glb	her	glb	1	elp	cnt	s.rnd	ent	1	hyp	opp	smp	her	glb	0	0	elp	atn	acu	ent	1	hyp	4-an	g
indernia procumbens	r	glb	her	glb	1	elp-s.orb	cnt	s.rnd	ent	1	hyp	opp	smp	her	glb	1	0	elp-ov	cnt	acu	ent	3	acr	4-an	
Aazus pumilus	r	glb	her	glb	1	b.ov	s.rnd	acu	ent	1	hyp	opp	smp	her	pub	1	0	ov	atn	obt	ent	1	cam		
Mecardonia procumbens	r	glb	her	glb	1	ov	cnt	obt	ent	1	hyp	opp		her	glb	1	0	b.ov	b.cnt	obt	ent	1		4-an	
Scoparia dulcis	r	scb	her	glb	1	ov c orb	cnt	acu	ent	1	hyp	opp		her	pub	1	0	ov b.ov	ent	acu	ent	1		4-an	S
Hygrophilla difformis	r	glb	her	glb	1	s.orb	s.rnd	s.rts	ent	1	hyp	opp	smp	her	pub	1		b.ov	cnt	obt	ent	1	cam	4-an 4-an	pi pi
eucas aspera	r	hrs	her	scb	1	obl-rec	b.cnt	rnd	ent	1	hyp	opp	smp	her	pub	1	0	n.elp	atn	acu	srt	1	cam		

Abbreviations: act: actinodromous; acm: acuminate; acu: acute; alt: alternate; ang: angular; atn: attenuate; aur: auriculate; b: broadly; bip: bipinnate; cam: camptodromous; cmp: compound; cnt: cuneate; crs: coriaceous; crd: cordate; cre: crenate; dnt: dentate; elp: elliptic; emg: emarginate; ent: antire; ep: epigeal; flb: flabellate; flc: falcate; glb: glabrous; her: herbaceous; hsp: hispid; hrs: hirsute; hyp: hyphodromous; irr: irregularly; lnc: lanceolate; lin: linear; min: minutely; muc: mucronate; n: narrowly; obl: oblong; obt: obtuse; obv: obovate; obq: oblique; opp: opposite; orb: orbicular; ov: ovate; plt: peltate; pm: palmately; prl: parallel; pub: pubescent; r: round; ren: reniform; rhm: rhomboidal; rnd: rounded; rtn: rectangular; rts: retuse; s: sub; scb: scabrous; sh: shallowly; sint: sinuately; smp: simple; sp: sparsely; spn: spiny; srt: serrate; str: strigose; tk: thick; tmn: tomentose; trn: truncate; wvy: wavy; 0: absent; 1: present; 3an: triangular; 4an: 4-angular; (species are arranged according to table 1)

- 5. First two leaves with apex emarginate or retuse; margin of subsequent leaves entire, first internode glabrous.......Amaranthaceae
- 5a. First two leaves with apex subrounded or obtuse; margin of subsequent leaves toothed, first internode hairy...........Chenopodiaceae
- 6. Venation of paracotyledons actinodromous; apex of
- first two leaves retuse..... ...Convolvulaceae [Evolvulus nummularius]
- 6a. Venation of paracotyledons hyphodromous; apex of first two leaves acute or obtuse......7
- 7. First two leaves elliptic or obovate oblong, base attenuate; length of first internode comparatively short (±1mm)..... ...Asteraceae

1	First two leaves ovate, base oblique or subrounded; ength of first internode comparatively long (5-	4a. 5.	Venation of first two leaves camptodromous5 First two leaves ovate							
	12mm) Solanaceae		Onagraceae [Ludwigia perennis]							
	to the families of Type II	5a.	First two leaves otherwise							
1.	Subsequent leaves simple2	6.	Venation of first two leaves acrodromous; first							
1a. 2.	Subsequent leaves compound6 Stipule ochreatePolygonaceae	0.	internode roundAsteraceae [exception Centipeda minima]							
2a.	Stipule linear	6a.	Venation of first two leaves camptodromous or							
3.	Venation of paracotyledons hyphodromous; first two leaves glabrous, venation hyphodromous; first	ou.	hyphodromous; first internode angular7 [exception <i>Lindernia procumbens</i>]							
	internode angular, glabrous, comparatively short (±1mm)	7.	Paracotyledons hairyBoraginaceae							
20		7a.	Paracotyledons glabrous8							
3a.	Venation of paracotyledons acrodromous or actinodromous; first two leaves hairy, venation actinodromous; first internode hairy, round, comparatively long (3.5-10mm)	8.	Hypocotyl reduced (± 1 mm); paracotyledons elliptic or ovate, comparatively smaller (1.5-2mm \times 1-1.5 mm)Scrophulariaceae							
4.	Paracotyledons oblong; apex of first two leaves acute; seedlings aromatic; latex and stellate hair presentEuphorbiaceae [Croton	8a.	Hypocotyl elongating; paracotyledons suborbicular or rectangular-oblong, comparatively larger (2.5-6mm ×2-4mm)9							
	bonplandianus]	9.	Apex of paracotyledons subretuse, margin of first							
4a.	Paracotyledons suborbicular; apex of first two		two leaves entire Acanthaceae [Hygrophila difformis]							
	leaves obtuse; seedlings not aromatic; latex and stellate hair absent5	9a .	Apex of paracotyledons rounded, margin of first two							
5.	Paracotyledons hirsute, primary veins 3; first two	7 a. 1	leaves serrateLamiaceae							
٥.	leaves ovate-rhomboid, base cuneate; first	Kos	to the families of Type IV							
	internode hirsuteMalvaceae [Sida	•	• •							
5a.	rhomboidea] Paracotyledons glabrous, primary veins 5; first two leaves suborbicular to broadly ovate, base subrounded; first internode pubescent		aracotyledons hairy, venation camptodromous or acrodromous; first two leaves ovate, base subtruncate or rounded, venation actinodromous, comparatively larger (3-7mm × 2-7 mm)							
	Sterculiaceae [Melochia corchorifolia]		Paracotyledons glabrous, venation hyphodromous;							
6.	Paracotyledons with venation actinodromous; venation of first two leaves camptodromous; first internode angular; apex of subsequent leaves not		first two leaves narrowly elliptic, base cuneate, venation hyphodromous, comparatively smaller (1-2mm × 1-2mm)Rubiaceae							
	emarginateFabaceae		Apex of paracotyledons retuse, venation							
6a.	Paracotyledons with venation hyphodromous; venation of first two leaves actinodromous; first internode round; apex of subsequent leaves emarginateOxalidaceae [Oxalis		camptodromous; first two leaves hirsute, base subtruncate, margin entire; subsequent leaves opposite decussate; first internode angular Urticaceae [Pouzolzia zeylanica]							
	corneculata]		Apex of paracotyledons subtruncate, venation							
Key	to the families of Type III		acrodromous; first two leaves scabrous, base rounded, margin serrate; subsequent leaves							
1.	Hypocotyl angularCaryophyllaceae		alternate; first internode round							
1a.	Hypocotyl round2	17	_							
2.	Seedlings with milky latex	_	to the genus/species of Type I							
2	Euphorbiaceae [Euphorbia hirta]	-	averaceae							
2a.	Seedlings without milky latex		aracotyledons sessile, apex acute; first two leaves sessile, margin spiny dentate							
3.	Margin of subsequent leaves entire4									
3a. 4.	Margin of subsequent leaves serrate or toothed		Paracotyledons petiolate, apex obtuse; first two leaves petiolate, margin pinnatisect							
	Venation of first two leaves hyphodromous		Fumaria indica							

Amaranthaceae

Key to the species under Amaranthus

- 1. First two leaves obovate, apex emarginate......
 Amaranthus gangeticus
- 1a. First two leaves ovate-elliptic, apex retuse......

 Amaranthus viridis

Chenopodiaceae

Key to the species under Chenopodium

- 1. First two leaves ovate-elliptic, apex subroundedChenopodium ambrosioides

Apiaceae

- 1. First two leaves trilobed, base subtruncate, margin entire, primary veins three; subsequent leaves compound..... *Oenanthe benghalensis*
- 1a. First two leaves not lobed, reniform, base subcordate, margin crenate, primary veins more than three; subsequent leaves simple....2

Asteraceae

Solanaceae

- 1. Hypocotyl hairy; paracotyledons ovate or ovatelanceolate, apex acute, comparatively larger (4-9 mm × 3-5 mm)......2

Key to the species under Physalis

- 1. First two leaves glabrous; first internode sparsely pubescent...... *Physalis minima*

Key to the genus/ species of Type II

Molluginaceae

Key to the species under Glinus

- 1. Paracotyledons ovate-suborbicular; first two leaves ovate-elliptic..... *Glinus oppositifolius*
- 1a. Paracotyledons obovate-suborbicular; first two leaves obovate-elliptic......Glinus lotoides

Polygonaceae

- 1. Apex of paracotyledons obtuse......2
- 1a. Apex of paracotyledons acute or rounded.....

 Persicaria
- 2. Paracotyledons sessile, linear-falcate; first two leaves linear, apex acute, venation hyphodromous...........Polygonum plebeium

Key to the species under Persicaria

- 1. Paracotyledons obtuse-suborbicular, venation camptodromous.........Persicaria hydropiper
- 1a. Paracotyledons linear, venation hyphodromous......

 Persicaria orientalis

Fabaceae

- 1. Paracotyledons falcate-oblong, base oblique; first two leaves simple, comparatively smaller (2.5-3mm × 3-3.5mm)... *Desmodium triflorum*

Key to the genus/ species of Type III

Caryophyllaceae

- 1a. Hypocotyl reduced (±1mm); paracotyledons ovateelliptic, apex subrounded; first two leaves obovate, base cuneate, apex subrounded.......Polycarpon prostratum

Amaranthaceae

Key to the species under Alternanthera

1. First two leaves oblanceolate, petiole longer (4-1a. First two leaves obovate, petiole reduced (1-1. Paracotyledons coriaceous, primary veins three, venation acrodromous..... Xanthium strumarium 1a. Paracotyledons herbaceous, primary veins one, venation hyphodromous or camptodromous......2 2. Venation of first two leaves hyphodromous; subsequent leaves spirally alternate..... Centipeda minima 2a. Venation of first two leaves acrodromous; subsequent leaves opposite decussate......3 3. Paracotyledons obovate-suborbicular, first two leaves with margin entire..... Eclipta prostrata 3a. Paracotyledons ovate, first two leaves with margin Key to the species under Ageratum conyzoides haustonianum Boraginaceae Paracotyledons hirsute; base of first two leaves cuneate, margin dentate..... Coldenia procumbens 1a. Paracotyledons pubescent; base of first two leaves rounded, margin irregularly wavy.....Heliptropium indicum Scrophulariaceae Paracotyledons ovate or broadly ovate......2 1a. Paracotyledons elliptic or elliptic-suborbicular (exceptions Lindernia ciliata, L. crustacea).....Lindernia Apex of paracotyledons acute; venation of first two leaves camptodromous......3

2a. Apex of paracotyledons obtuse; venation of first

Base of paracotyledons subrounded; apex of first

leaves acute.....Scopariadulcis

First two leaves oblong-spathulate, hirsute, margin

spiny dentate.....Lindernia ciliata

glabrous, margin entire or inconspicuously

crenate.....2

1a. First two leaves ovate, elliptic or ovate-elliptic,

3a. Base of paracotyledons cuneate; apex of first two

Key to the species under Lindernia

- 2. First two leaves with three primary veins, venation acrodromous............Lindernia procumbens

- 3a. Paracotyledons ovate, base subtruncate, apex obtuse................Lindernia crustacea
- 4. Apex of paracotyledons acute, first two leaves petiolate.....Lindernia nummulariifolia
- 4a. Apex of paracotyledons subrounded; first two leaves sessile........Lindernia parviflora

Lamiaceae

- 1a. Hypocotyl comparatively reduced (2-3mm); paracotyledons suborbicular; first two leaves ovate, apex obtuse; first internode comparatively shorter (3-5mm)...... Salvia plebeia

Key to the genus/ species of Type IV

Euphorbiaceae

Rubiaceae

- 1. Paracotyledons elliptic, apex obtuse; base of first two leaves cuneate......Dentella repens

RESULTS AND DISCUSSION

The artificial key revealed that all the seedlings are of phanerocotylar epigeal types. In the artificial key, the sixty taxa from twenty four families of Magnoliopsida have been separated into four types based on naturally adapted characters such as phyllotaxy and presence of stipules of the first two leaves. Each type contains many families with exclusively all studied taxa but some types contain taxa not confined to it exclusively but to other types also. Thus, type I consists of families such as Papaveraceae, Ranunculaceae, Apiaceae, Chenopodiaceae, Convolvulaceae, Solanaceae, genus

Amaranthus of Amaranthaceae, genera Grangea and Gnaphalium of Asteraceae. Type II is represented by the families Polygonaceae, Molluginaceae, Malvaceae, Sterculiaceae, Fabaceae, Oxalidaceae and genus Croton of Euphorbiaceae. Type III contained families like Caryophyllaceae, Portulacaceae, Onagraceae, Boraginaceae, Scrophulariaceae, Acanthaceae, Lamiaceae, and the genus Euphorbia of Euphorbiaceae, the genera Achyranthus and Alternanthera of Amaranthaceae, and four genera of Asteraceae. At last Rubiaceaea, Urticaceae and two genera of Euphorbiaceae (Acalypha and Chrozophora) belonged to seedling type IV.

This classification is totally based on seedling morphological traits and showsed somehomology with other traditional system of plant classifications. For example, under type I, Amaranthaceae and Chenopodiaceae share some common characters such as narrowly oblong paracotyledons with obtuse apex and angular first internode. They also belong to the order Caryophyllales in Takhtajan's system (1997). Similarly, Malvaceae and Sterculiaceae under type II display common juvenile traits like simple subsequent leaves, suborbicular paracotyledons, obtuse apex of first two leaves they remain together in order Malvales under subclass Dilleniidae of Takhtajan's system and they differ from Fabaceae and Oxalidaceae having compound subsequent leaves. The latter two are belonging to subclass Rosidae. Under type III, Scrophulariaceae, Acanthaceae and Lamiaceae (subclass Lamiidae of Takhtajan, 1997) remain associated together having round hypocotyl, margin of subsequent leaves serrate or toothed, venation of first two leaves camptodromous or hyphodromous, angular first internode. Similarly, Urticaceae and Euphorbiaceae show close affinity based on hairy paracotyledons with camptodromous or acrodromous venation; first two leaves ovate with base subtruncate or rounded and venation actinodromous supporting quite parallel inclusion of them under subclass Dillenidae in Takhtajan's system (1997).

Juvenile characters have also supported few other botanical disciplines such as pollen morphology, cytology, phytochemistry, etc. of the studied taxa. Fatinah et al. (2012) worked on phylogeny of six members of Amaranthaceae using RAPD and showed that Achyranthes and Alternanthera tend to stay together while Amaranthus separated out from them. From juvenile traits, it has been seen that Amaranthus having sub-opposite to alternate exstipulate first two leaves belonging to type I seedling while the other two taxa having opposite, exstipulate first two leaves

represented type III, thus supporting their phylogenetic correlation based on RAPD. A study on the pollen grains of a few *Chenopodium* spp. by Pinar and Inceoglu (1999) showed that C. album and C. ambrosioides share similar radial symmetrical, isopolar, pantopolyporate spheroidal pollen grains with scabrate ornamentation. In seedling morphology, these two taxa also share some similar characters such as first two leaves coriaceous, apex subrounded or obtuse; margin of subsequent leaves toothed and first internode hairy indicating the similarities between these two taxa. Rahman et al, (2013) investigated stomata and trichome characters of 36 species of Asteraceae and showed that while Ageratum, Grangea and Gnaphalium share similar characters such as anomocytic stomata and nonglandular multicellular trichomes but Xanthium displays anisocytic stomata and non-glandular unicellular trichomes. Our seedling study contradicts with their findings in such a way that Grangea and Gnaphalium represent seedling type I while Ageratum and Xanthium belonging to type III, thus partially supporting the above study.

Although this work was considered keeping in mind about the identification of weeds in seedling stage which is crucial for eradication and creating an opportunity towards integrated weed management (Chomas et al. 2001, Parkinson et al. 2013) but after the discussion it is clear that study of weeds at juvenile stage has some taxonomic values too. The morphological markers used for the identification of the taxa are useful for delimitation of the taxa into different taxonomic groups showing an insight about the usefulness of juvenile features. The traits are highly conservative and hence the key is viable independent of habitat, climate or soil nature. The comparison with an accepted existing system and a few other botanical disciplines gives new ideas about plant systematic which are vital in many a ways.

Furthermore, the most applied outcome of this work is weed management through seedlings. Since, number of seed production, seed production rate, seed viability and resistance to environment are higher in weeds compared to crops, vigorous germination of seeds occur during pre- or post- harvest periods establishing abundant seedlings. Even occurrence of flowering can be observed in these weeds at seedling stage indicating their partly ephemeral behaviour. This makes it a lot harder for their total eradication from the field because they immediately disperse enumerable seeds. However, eradication at the seedling stage minimizes the chance of further weed dispersal by limiting their life cycle before flowering.

Thus, proper identification of weeds at seedling stage may reduce the chemical herbicides proving it to be more economic and eco-friendly.

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